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Evaluation of grain yield based on digital images of rice canopy

Kailou Liu^{1,2*} , Yazhen Li¹, Tianfu Han², Xichu Yu¹, Huicai Ye¹, Huiwen Hu¹ and Zhihua Hu¹

Abstract

Background: Rice canopy changes are associated with changes in the red light (R), green light (G), and blue light (B) value parameters of digital images. To rapidly diagnose the responses of rice to nitrogen (N) fertilizer application and planting density, a simple model based on digital images was developed for predicting and evaluating rice yield.

Results: N application rate and planting density had significant effects on rice yield. Rice yield first increased and then decreased with increasing of N rates, while the rice yield always increased significantly with increasing planting density. The normalized redness intensity (NRI), normalized greenness intensity (NGI), and normalized blueness intensity (NBI) values of the rice canopy varied among stages; however, they were primarily affected by N fertilizer rates, while planting density had no significant effects. Furthermore, the significant relationships of grain yield with NRI and NBI at the late filling stage could be fitted by quadratic equations, but there was no significant relationship observed between grain yield and NGI across all stages. In addition, a field validation experiment showed that the predicted yield based on the fitted quadratic equations was consistent with the measured yield.

Conclusion: The NRI, NGI, and NBI values of rice canopy were mainly affected by N fertilizer rates, while the planting density had no significant effect. The significant relationships between grain yield with NRI and NBI at the late filling stage could be fitted by quadratic equations. Therefore, the canopy NRI and NBI at the late filling stage as measured by digital photography could be used to predict grain yield in southern China.

Keywords: Digital camera, Rice canopy, Yield prediction, Late filling stage

Background

As one of the most important food crops in the world, rice feeds half of the global population, especially in East Asia and Southeast Asia [1, 2]. Most rice-dependent populations are currently in developing countries such as China, India, and Thailand [3, 4]. With increasing food demand in these and other regions, advanced technological innovations in rice production will be critical to stabilizing food security.

Rice yield is typically quantified after harvesting and drying. This process requires time, power, and resources. Therefore, the rapid and accurate estimation of rice yield

has become an important part of rice production technology research. With the development of remote sensing technology, more remote sensing tools have been applied to agricultural monitoring. Use of terrestrial hyperspectral remote sensing [5, 6] and satellite and aerial imagery [7–9] to conduct rice yield assessment have been reported. At present, the main methods for assessing rice yield primarily include spectroscopic diagnosis [9] and remote sensing evaluation [10], but these methods require the purchase of expensive spectrometers or satellite remote sensing images. Additionally, these methods are complex and give uncertain results [11], which makes them difficult to promote and apply.

However, owing to recent price decreases and integration with smartphones, digital cameras are now widely used [12]. Since the 1990s, digital cameras have been used as one of the most convenient tools for remote sensing of the visible light spectrum (in which R, G, and B represent the gray values from the red, green, and blue

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channels) in agricultural information monitoring, such as automatic classification of agricultural products [13], weed identification, and pest monitoring [14], among other applications. Kawashima et al. [15] reported manual selection of a complete blade for crop extraction (RB)/(R+B) to estimate the leaf chlorophyll content. Adamsen et al. [16] used a digital camera to vertically capture images of wheat canopies, and cut out images representing 1 m² to calculate parameters such as G/R. That study showed that G/R and the normalized difference vegetation index (NDVI) were significantly related to the chlorophyll metric, which was shown by soil-plant analyses development; (SPAD) value. Rorie et al. [17] used a digital camera to capture images of corn leaves under an active light source and calculated dark green color index (DGCI) after correction with a reference color, with results showing a strong correlation between nitrogen (N) content and yield of leaves. Li et al. [18] vertically photographed wheat canopies and segmented the images using soil adjustment vegetation index (SAVI) Green > 0 and found that the extracted coverage (Canopy Cover) positively correlated with the leaf area index (LAI), plant N content, and aboveground biomass. Previous studies in rice showed that leaf color measured with digital cameras can characterize N and chlorophyll content in rice [19, 20]. In addition, green-channel minus red (GMR) can further distinguish the N diagnosis of *indica* and *japonica* rice varieties [21]. In summary, the R, G,

and B values of digital photos are closely related to nutrient content in rice [18, 20].

The current applications for digital cameras in rice and other crops are limited to nutritional diagnosis, and only a few studies have reported the evaluation of rice yield with digital cameras [22]. Using digital cameras for predicting rice yields can quickly assess rice production and provide decision-making basis for rice farmers and for governments. In this study, a field experiment was conducted which included different N fertilizer application rates and planting densities, and the color images of the rice canopy in the key growth periods were measured, including values for R, G, and B and the normalized redness intensity (NRI), normalized greenness intensity (NGI), and normalized blueness intensity (NBI). Moreover, the fitted regressions between grain yield with NRI, NGI, and NBI values were established. Field verification was also conducted to evaluate the practicality of the fitted regression.

Results

Rice yield

N rates and planting density both had significant effects on rice yield (Fig. 1). Among all treatments, grain yield was highest in the N2D4 treatment with 8.34×10^3 kg ha⁻¹, while yield was the lowest in N0D1 treatment (5.01×10^3 kg ha⁻¹). Compared with the no N application (N0), the grain yields were higher in the

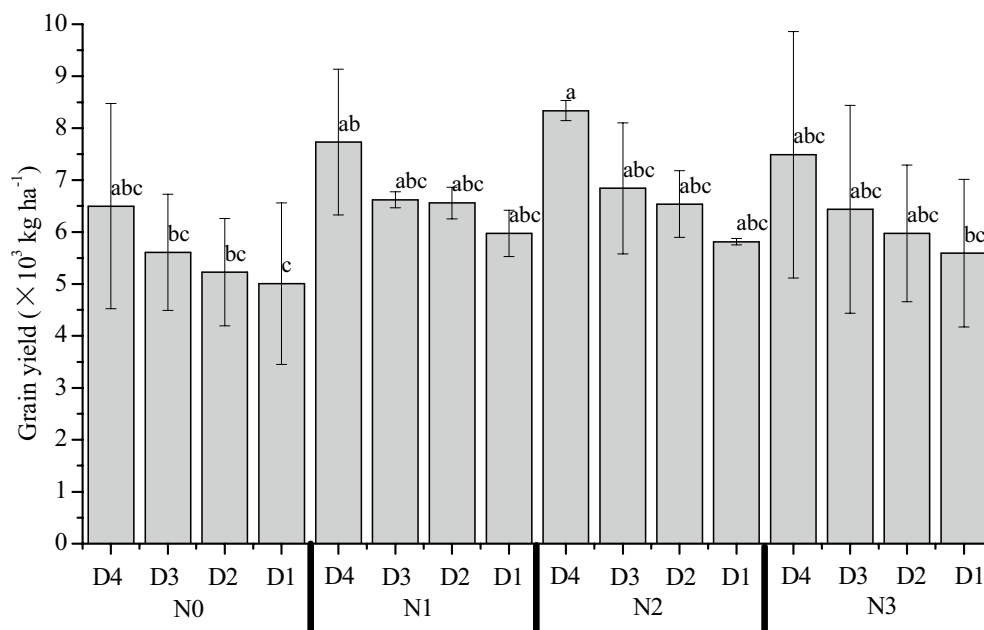


Fig. 1 Rice yields of different treatments with varied N and D. Error bar indicates SD. Different lowercase letters indicate significant differences ($p < 0.05$). N represents nitrogen, and D represents planting density

Table 1 The fitted regression between nitrogen (N) rates (x) and grain yield (y)

Planting densities	Fitted equations	R ²	p
D4	$y = -0.0669x^2 + 20.243x + 6473.3$	0.8636	0.42793
D3	$y = -0.0475x^2 + 14.651x + 5601.5$	0.9629	0.16541
D2	$y = -0.0757x^2 + 20.504x + 5222.3$	0.9917	0.11260
D1	$y = -0.0491x^2 + 13.566x + 5008.5$	0.9939	0.17084

D represents planting density

Table 2 The fitted regression between planting densities (x) and grain yield (y)

N rates	Fitted equations	R ²	p
N0	$y = 8107x + 3153.6$	0.9101	0.06309
N1	$y = 8881x + 4056.9$	0.8804	0.17204
N2	$y = 13119x + 2946.8$	0.9178	9.56E-04
N3	$y = 10260x + 3294.9$	0.9387	0.03314

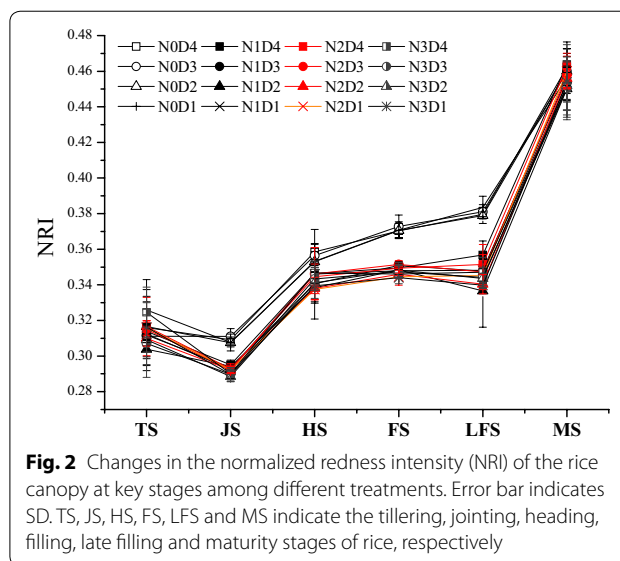
N represents nitrogen

Table 3 Comparative analysis of rice yield under nitrogen (N) and density treatments

Treatments	Yield ($\times 10^3$ kg ha ⁻¹)	Treatments	Yield ($\times 10^3$ kg ha ⁻¹)
N0	5.59 ± 0.67 b	D4	7.51 ± 0.76 A
N1	6.72 ± 0.73 ab	D3	6.38 ± 0.54 B
N2	6.88 ± 1.06 a	D2	6.08 ± 0.63 B
N3	6.37 ± 0.82 ab	D1	5.60 ± 0.42 B

Different lowercase letters indicate significant differences among nitrogen (N) fertilizer treatments ($p < 0.05$). Different uppercase letters indicate significant differences among different planting density treatments ($p < 0.05$)

N fertilizer application treatments, but the yields were not increased when N exceeded 180 kg ha⁻¹ (N2). There was no significant relationship between N rate and grain yield under the same planting density, as demonstrated through the fitted regressions (Table 1). The fitted regressions showed that grain yield could be significantly improved by increasing planting densities under the same N rate (Table 2). Furthermore, these equations also showed that the growth rate of rice yield was higher in the N2 and N3 treatments than in the N0 and N1 treatments. The yield of the N2 treatment increased by 23.22% over that of the N0 treatment, but the yield of the N1 and N3 treatments did not differ significantly than that of the N0 treatment (Table 3). Increasing rice planting density can significantly improve grain yield. The highest yield was observed in the D4 treatment, which was increased by 34.27% over that of the D1 treatment. However, the yields of D2 and



D3 treatments were not significantly increased over that of the D1 treatment.

Changes in the NRI, NGI, and NBI values of the rice canopy

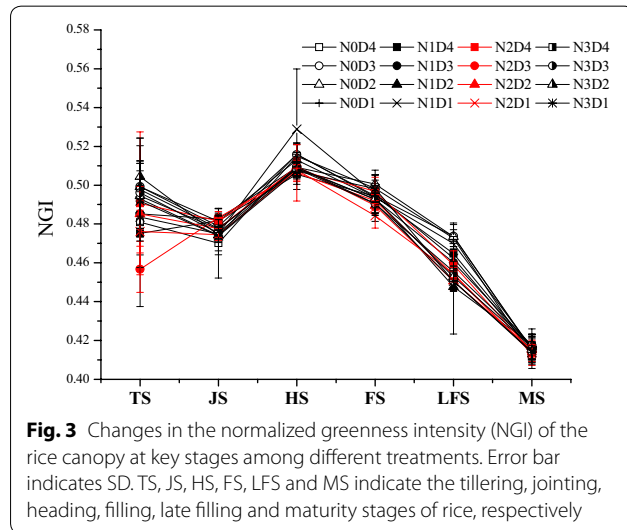
The NRI values among all treatments gradually increased with the development of rice growth, which can be roughly divided into three periods (Fig. 2): (1) the gradually increasing period was from the tillering to heading stage, (2) the stable period was from the heading to late filling stage, and (3) the rapidly increasing period was from the late filling to maturity stage. NRI in the key growth stages of rice was primarily regulated by the amount of N fertilizer, and different planting densities have little effect (Table 3). Across different growth periods, the NRI value in the N0 treatment was the highest among the most key stages, although different N rates also caused a significant difference in NRI values. Compared with the no N fertilizer treatment (Table 4), the NRI values in the N1 treatment were reduced by 4.86%, 3.51%, and 5.91% in the heading, filling, and late filling stages, respectively, while those in the N2 treatment decreased by 5.31%, 3.88%, and 6.27%, and those of the N3 treatment decreased by 6.25%, 3.57%, and 6.65% across these same stages.

The NGI values of all treatments gradually decreased with increasing rice growth, and these can be divided into three general periods as well (Fig. 3): (1) a stable period was from the tillering to the jointing stage, (2) a gradual increase period was from the jointing to the heading stage, and (3) a rapid reduction period was from the heading to the maturity stage. Similar to trends for NRI, NGI during the key growth stages of rice could be affected by the N rates, but different planting densities

Table 4 Comparative analysis of the normalized redness intensity (NRI) in rice canopy under nitrogen (N) fertilizer and density treatments

Treatments	TS	JS	HS	FS	LFS	MS
N0	0.317 ± 0.006a	0.309 ± 0.002a	0.355 ± 0.003a	0.371 ± 0.001a	0.381 ± 0.002a	0.460 ± 0.002a
N1	0.310 ± 0.004a	0.294 ± 0.001b	0.343 ± 0.004b	0.349 ± 0.001b	0.347 ± 0.008b	0.456 ± 0.006a
N2	0.314 ± 0.003a	0.292 ± 0.000b	0.342 ± 0.004b	0.348 ± 0.003b	0.346 ± 0.005b	0.459 ± 0.003a
N3	0.314 ± 0.008a	0.290 ± 0.001b	0.343 ± 0.003b	0.346 ± 0.002b	0.344 ± 0.003b	0.452 ± 0.002a
D4	0.322 ± 0.005A	0.297 ± 0.008A	0.347 ± 0.006A	0.353 ± 0.011A	0.356 ± 0.014A	0.456 ± 0.006A
D3	0.312 ± 0.003A	0.297 ± 0.010A	0.345 ± 0.007A	0.355 ± 0.012A	0.353 ± 0.017A	0.454 ± 0.004A
D2	0.308 ± 0.005A	0.296 ± 0.008A	0.340 ± 0.007A	0.352 ± 0.012A	0.349 ± 0.020A	0.454 ± 0.004A
D1	0.316 ± 0.006A	0.296 ± 0.008A	0.345 ± 0.007A	0.352 ± 0.013A	0.354 ± 0.020A	0.459 ± 0.003A

TS, JS, HS, FS, LFS and MS indicate the tillering, jointing, heading, filling, late filling and maturity stages of rice, respectively. Different lowercase letters indicate significant differences among nitrogen (N) fertilizer treatments ($p < 0.05$). Different uppercase letters indicate significant differences among planting density treatments ($p < 0.05$)



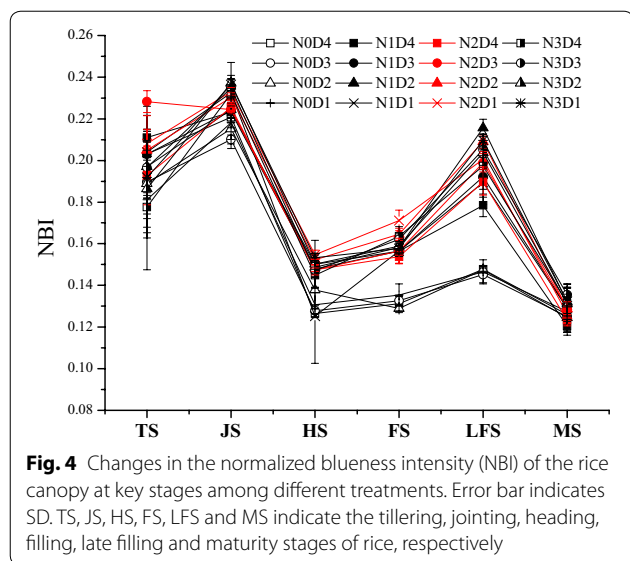
had little effect (Table 4). Among four N rates, the NGI value of the tillering stage was the highest in the N3 treatment. However, in the late filling stage, the NGI values for the N fertilizer treatments were significantly lower than that of the N0 treatment (Table 5). The NGI values in the N1, N2, and N3 treatments decreased below that of the N0 treatment by 2.90%, 3.82%, and 4.26%, respectively.

Across all treatments, NBI values gradually decreased along with rice growth, and could be roughly divided into four periods, which include: (1) the gradual increase period was from the tillering to the jointing stage, (2) the rapid reduction period was from the jointing to the heading stage, (3) the increase period was from the heading to the late filling stage, and (4) the stabilization period was from the late filling to the maturity stage (Fig. 4). Like NRI and NGI, the NBI values in the key growth stages of rice were also affected by N rates (Table 5), while the different

Table 5 Comparative analysis of the normalized greenness intensity (NGI) in rice canopy under nitrogen (N) fertilizer and density treatments

Treatments	TS	JS	HS	FS	LFS	MS
N0	0.492 ± 0.008ab	0.475 ± 0.004a	0.514 ± 0.003a	0.497 ± 0.003a	0.472 ± 0.002a	0.415 ± 0.002a
N1	0.486 ± 0.010b	0.480 ± 0.004a	0.514 ± 0.010a	0.494 ± 0.001a	0.459 ± 0.008b	0.416 ± 0.001a
N2	0.477 ± 0.015ab	0.479 ± 0.004a	0.508 ± 0.001a	0.491 ± 0.005a	0.454 ± 0.003b	0.414 ± 0.001a
N3	0.498 ± 0.005a	0.475 ± 0.002a	0.508 ± 0.001a	0.493 ± 0.004a	0.452 ± 0.002b	0.416 ± 0.002a
D4	0.479 ± 0.010A	0.478 ± 0.005A	0.509 ± 0.004A	0.496 ± 0.003A	0.462 ± 0.008A	0.417 ± 0.002A
D3	0.481 ± 0.019A	0.480 ± 0.004A	0.510 ± 0.004A	0.492 ± 0.002A	0.458 ± 0.010A	0.417 ± 0.002A
D2	0.494 ± 0.008A	0.479 ± 0.003A	0.512 ± 0.002A	0.496 ± 0.005A	0.456 ± 0.012A	0.417 ± 0.001A
D1	0.484 ± 0.008A	0.474 ± 0.000A	0.515 ± 0.010A	0.492 ± 0.006A	0.458 ± 0.009A	0.415 ± 0.001A

TS, JS, HS, FS, LFS and MS indicate the tillering, jointing, heading, filling, late filling and maturity stages of rice, respectively. Different lowercase letters indicate significant differences among nitrogen (N) fertilizer treatments ($p < 0.05$); different uppercase letters indicate significant differences among planting density treatments ($p < 0.05$)



planting densities had no significant effect. There was no significant difference for NBI in the maturity stage across different treatments. In the tillering stage, the NBI values of the N1 and N2 treatments were significantly higher than for the N0 and N3 treatments, but in all other stages, the NBI values in the N0 treatment were the lowest. However, compared with the N0 treatment, the NBI values were significantly higher in the N fertilizer (N1, N2, and N3) treatments (Table 6). In the jointing, heading, filling, and late filling stages, the NBI values of the N1 treatment decreased below that of the N0 treatment by 4.62%, 9.37%, 18.81%, and 32.18%, respectively. The NBI values of the N2 treatments were reduced by 5.68%, 15.12%, 22.30%, and 35.99%, respectively, and the NBI values of the N3 treatments were reduced by 8.88%, 13.90%, 21.59%, and 39.03%, respectively.

Relationships between rice yield and NRI, NBI, and NGI

Across different key growth stages of rice, the relationships between yield and NRI or NBI of rice canopy digital images at the late filling stage were fitted by quadratic curves (Table 7), with $y = -3437.4x^2 + 2465.5x - 434.79$ ($R^2 = 0.4853, p < 0.05$); and $y = -1198.9x^2 + 436.5x - 32.612$ ($R^2 = 0.4122, p < 0.05$), respectively. However, there was no significant relationship between NGI and yield.

Model verification

In field validation experiments, the predicted yields were attained through the fitted equations with NRI and NBI at the late filling stage (Fig. 5). Concurrence between the simulated and measured values was high, with R^2 of 0.4592 and 0.7074 and RMSE of 0.5489 and 0.4010, respectively, with average RE values of -0.024 and -0.028 . Therefore, the NRI and NBI at the late filling stage can be used to better predict grain yield.

Discussion

The application of digital cameras and image processing technology can be used to quickly obtain crop canopy data, and this method is cheaper and easier to operate and implement than other methods, such as hyperspectral remote sensing and satellite imagery. The rapid increase in the use of smartphones (with a self-contained camera function) can enable this technology to meet huge potential applications for rice planting. In this study, with increases in rice growth (from the tillering to the maturity stage), NRI values increased gradually, while NGI and NBI peaked at the heading and jointing stages, respectively, and then gradually decreased. This may be due to growth and metabolism consuming a great deal of energy, because NRI is directly related to available energy [23, 24]. Moreover, NRI, NGI, and NBI in key growth stages of rice were

Table 6 Comparative analysis of the normalized blueness intensity (NBI) in rice canopy under nitrogen (N) fertilizer and density treatments

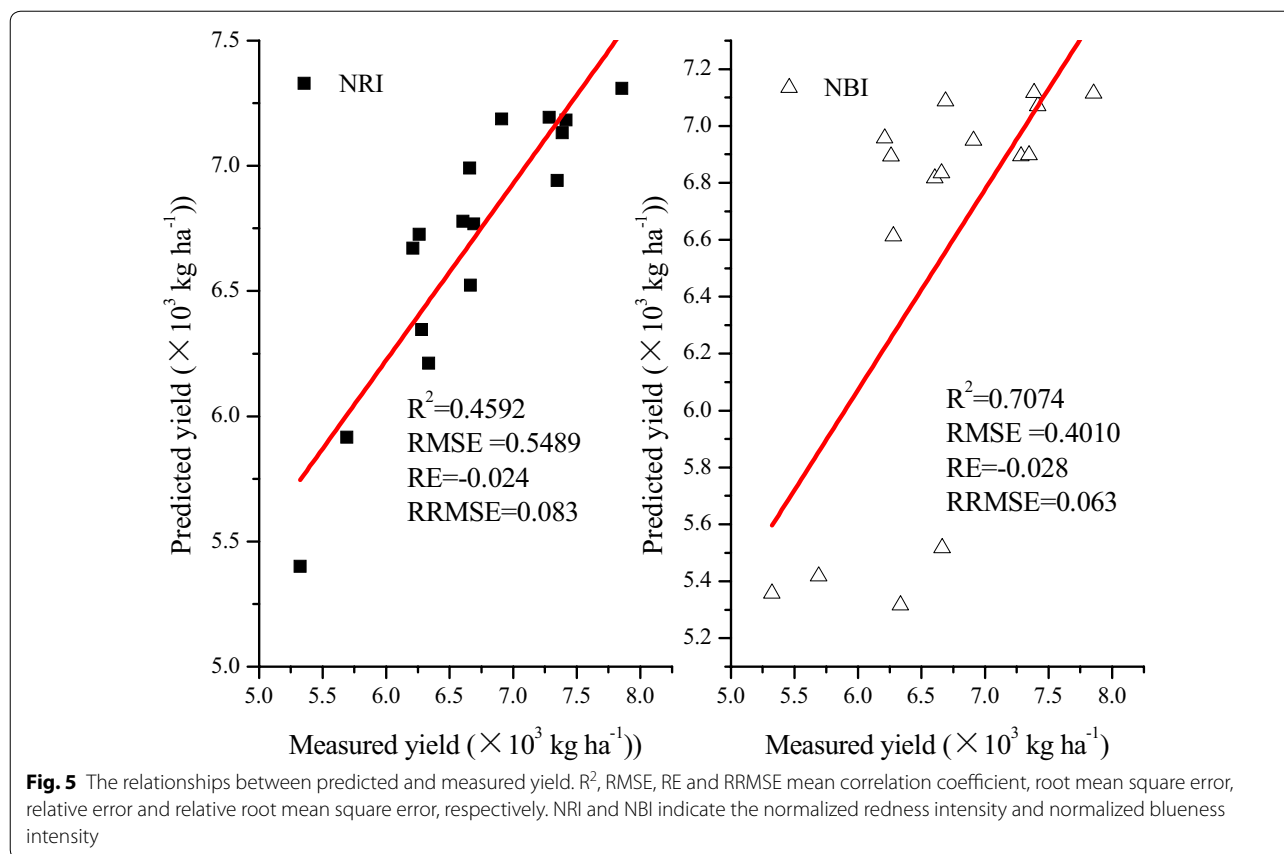
Treatments	TS	JS	HS	FS	LFS	MS
N0	0.191 ± 0.009 b	0.216 ± 0.005c	0.131 ± 0.005b	0.132 ± 0.003b	0.147 ± 0.001b	0.126 ± 0.001a
N1	0.204 ± 0.006 a	0.226 ± 0.004b	0.143 ± 0.012a	0.157 ± 0.001a	0.194 ± 0.016a	0.128 ± 0.005a
N2	0.209 ± 0.015 a	0.228 ± 0.004b	0.150 ± 0.004a	0.162 ± 0.008a	0.200 ± 0.008a	0.127 ± 0.003a
N3	0.188 ± 0.008b	0.235 ± 0.002a	0.149 ± 0.004a	0.161 ± 0.003a	0.204 ± 0.005a	0.133 ± 0.002a
D4	0.199 ± 0.014A	0.226 ± 0.005A	0.144 ± 0.010A	0.151 ± 0.014A	0.182 ± 0.022A	0.126 ± 0.005A
D3	0.207 ± 0.017A	0.223 ± 0.010A	0.145 ± 0.010A	0.153 ± 0.013A	0.188 ± 0.027A	0.128 ± 0.004A
D2	0.197 ± 0.008A	0.225 ± 0.009A	0.148 ± 0.006A	0.152 ± 0.016A	0.195 ± 0.031A	0.129 ± 0.003A
D1	0.200 ± 0.012A	0.230 ± 0.008A	0.141 ± 0.015A	0.156 ± 0.015A	0.188 ± 0.028A	0.125 ± 0.003A

TS, JS, HS, FS, LFS and MS indicate the tillering, jointing, heading, filling, late filling and maturity stages of rice, respectively. Different lowercase letters indicate significant differences among nitrogen (N) fertilizer treatments ($p < 0.05$); different uppercase letters indicate significant differences among planting density treatments ($p < 0.05$)

Table 7 Fitted equations for the normalized redness intensity (NRI), normalized greenness intensity (NGI), and normalized blueness intensity (NBI) values (x) and yield (y) across different growth stages of rice

Stages	NRI		NGI		NBI				
	Regression equations	R ²	p	Regression equations	R ²	p			
TS	$y = -2E + 0.6x^2 + 1E + 0.6x - 188418$	0.0109	0.9311	$y = -119334x^2 + 96588x - 12308$	0.0597	0.67032	$y = 245939x^2 - 80957x + 12745$	0.0664	0.63984
PS	$y = -9E + 0.6x^2 + 5E + 0.6x - 783277$	0.2824	0.11566	$y = 2E + 0.7x^2 - 2E + 0.7x + 4E + 0.6$	0.3632	0.05322	$y = -8E + 0.6x^2 + 4E + 0.6x - 403687$	0.3531	0.05894
HS	$y = -8E + 0.6x^2 + 6E + 0.6x - 976732$	0.1738	0.28902	$y = 7E + 0.6x^2 - 7E + 0.6x + 2E + 0.6$	0.226	0.18915	$y = -4E + 0.6x^2 + 1E + 0.6x - 73035$	0.1926	0.24896
FS	$y = -9E + 0.6x^2 + 6E + 0.6x - 1E + 0.6$	0.4148	0.03074	$y = -250780x^2 + 158991x - 10957$	0.1883	0.46575	$y = -2E + 0.6x^2 + 755848x - 51130$	0.293	0.10504
LFS	$y = -3E + 0.6x^2 + 2E + 0.6x - 434790$	0.4853	0.01333	$y = -158882x^2 + 99109x - 5580.9$	0.2439	0.10197	$y = -1E + 0.6x^2 + 436500x - 32612$	0.4122	0.03163

TS, JS, HS, FS, LFS and MS indicate the tillering, jointing, heading, filling, late filling and maturity stages of rice, respectively



mainly regulated by N fertilizer application, rather than by planting density. Compared with no N application, the NRI values of the heading stage in the N fertilizer treatments were significantly lower, but the NBI values were significantly higher. The R, G, B values could change as a result of leaf yellowing and senescence, which can be caused by insufficient nutrient supply [25, 26].

The R, G, and B values of plant canopies can reflect nutrient uptake, especially that of N content and uptake by crops [17–19]. Previous studies have shown that for rice, wheat, corn, and cotton, the R, G, B values obtained by a camera can reflect the N uptake capacity of plants, and can be further used to diagnose N deficiency and assess biomass and grain yield [18, 20, 22]. The current study also found that the relationship between NRI or NBI and grain yield can be fitted by a quadratic curve across all key rice growth stages. This result was the same as other studies, which all suggested that grain yield was closely related to NRI and NBI [27, 28]. In these field experiments, the rainfall and temperature that occurred in the second year (2017) of the verification experiments were similar to those of the first year (2016). The simulated values of rice yield obtained by fitting equations were highly consistent with the measured values, with high estimation accuracy and low average relative error. NRI and NBI at the late filling stage can thus be used

to better predict grain yield. However, rice yield could change as a result of the interaction of complex factors [29, 30], such as unusual precipitation and temperature, which can cause rice lodging and disease [31–33]. Therefore, actual rice yield may differ from the yield predicted by NRI and NBI at the late filling stage, so these predictions should be used cautiously.

Conclusion

Metrics of growth in rice canopy across different growth stages can be measured and assessed by changes to the R, G, and B parameters of digital images. The NRI, NGI, and NBI of the rice canopy varied significantly across different N fertilizer rates, though the measured rice yield varied depending on both N fertilizer rate and planting density. NRI and NBI at the late filling stage could be used to predict grain yield through using the fitted quadratic curve equations, and these results were upheld by the field validation experiment.

Methods

Site description

The field experiment was conducted at Yanjia Village, Zhanggong Town, Jinxian County, Nanchang City, Jiangxi Province, China (116°24'E, 28°15'30"N). This area

experiences a mid-subtropical monsoon climate, with an average annual rainfall of 1537 mm, annual evaporation of 1150 mm, annual average temperature of 18.1 °C, with average temperatures in the coldest month (January) and the hottest month (July) of 4.6 °C and 29.1 °C, respectively. In the experimental years of the study (2016 and 2017), rainfall primarily occurred in March, April, May, and June (Fig. 6). The proportion of rainfall received in this season was 61.14% for 2016 and 52.34% for 2017. The rice growing season (from July to November) experiences higher temperature and lower rainfall. Average temperatures from July to November were 23.49 °C and 23.49 °C in 2016 and 2017, respectively, and the total rainfalls were 435.40 mm and 812.90 mm. The altitude is 25–30 m, a typical low hilly area. The soil type is paddy soil developed by Quaternary red clay. Soil pH is 6.9, the organic carbon is 16.22 g kg⁻¹, total N is 0.95 g kg⁻¹, total phosphorus is 1.02 g kg⁻¹, total potassium is 15.41 g kg⁻¹, and alkaline N is 143.70 mg kg⁻¹; available phosphorus is 10.30 mg kg⁻¹, and available potassium is 125.10 mg kg⁻¹.

Experimental design

In this study, the field experiment was conducted in 2016 and 2017 for model establishment and validation, respectively. The main treatment was N fertilizer rates (0, 135, 180, and 225 kg ha⁻¹ N) and the secondary treatment was density levels (0.21, 0.27, 0.33 and 0.39 million plants ha⁻¹). Each treatment was three replications, and main area was

9 × 4.5 = 40.5 m² (the sub-area was 2.2 × 4.5 = 9.9 m²), and the random area group was arranged.

Chemical fertilizer was applied to treatments (Table 8). The application ratios for N fertilizer were 40%, 30%, and 30% in basal, tiller, and panicle fertilizers, respectively. All treatments (including the no N fertilizer treatment) received application of 60 kg ha⁻¹ P₂O₅ with calcium magnesium phosphorus (12.5% P₂O₅) and 225 kg ha⁻¹ K₂O with potassium chloride (60% K₂O). All phosphorus and 50% of the potassium fertilizer were used as base fertilizer, while the remaining 50% potassium fertilizer was applied as panicle fertilizer. The application time for basal, tiller, and panicle fertilizers was 1 day before transplanting rice, 10 days and 45 days after transplanting rice, respectively.

The rice variety was ‘Zhengcheng 456’, which was sown on 25th June, transplanted on 24th July, harvested on 5th November. Water, weeds, insects, and disease were controlled as required to avoid yield loss.

Measurement index

- 1) Rice yield determination

Matured rice plants in each plot were harvested for threshing and measured for standard yield after drying (water content was 13.5%).

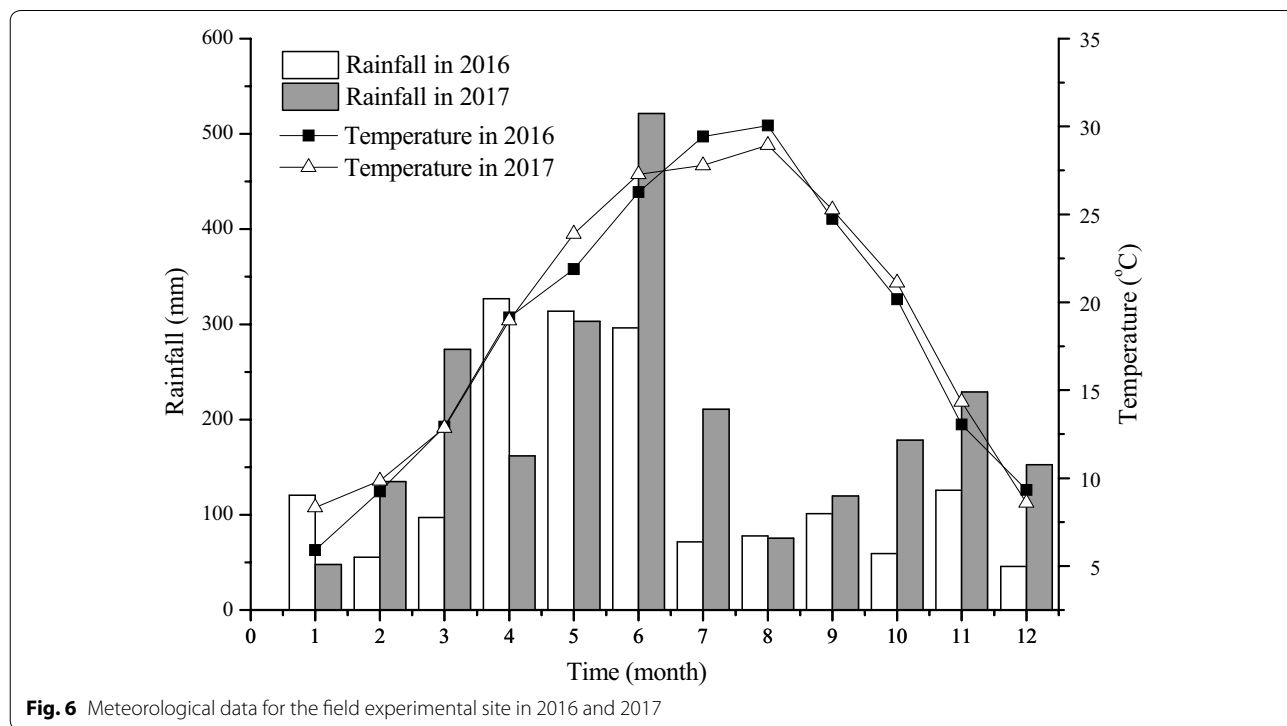


Table 8 The nitrogen (N) fertilizer rates and planting density in different treatments

Treatments	N fertilizer rates (kg ha ⁻¹)	Planting densities (million plants ha ⁻¹)	Plant spacing (× 10 ⁻² m)	Line spacing (× 10 ⁻² m)
N0D1	0	0.21	20	23.8
N0D2	0	0.27	20	18.5
N0D3	0	0.33	20	15.2
N0D4	0	0.39	20	12.8
N1D1	135	0.21	20	23.8
N1D2	135	0.27	20	18.5
N1D3	135	0.33	20	15.2
N1D4	135	0.39	20	12.8
N2D1	180	0.21	20	23.8
N2D2	180	0.27	20	18.5
N2D3	180	0.33	20	15.2
N2D4	180	0.39	20	12.8
N3D1	225	0.21	20	23.8
N3D2	225	0.27	20	18.5
N3D3	225	0.33	20	15.2
N3D4	225	0.39	20	12.8

N represents nitrogen; and D represents planting density

2) Photographing rice canopy

In the tillering, jointing, heading, filling, late filling, and maturity stages, images of the rice canopy of rice in 2016 and 2017 (Additional file 1: Figs. S1 and S2) were obtained in the field with a Canon IXUS140 digital camera following established methods [34]. Crops were photographed at a vertical height of 1.2 m from the ground (about 1 m from the rice canopy) and at a 60° angle to the ground. A 15 × 5 cm white plastic plate was used as the background for shooting in the camera's automatic white balance mode. The image resolution was 1280 × 960, and the camera's image of the ground rice canopy range was approximately 1.2 m × 1 m trapezoids. Digital images were transferred to the computer in JPEG format.

The image was processed using Adobe Photoshop. "Color selection" was used to select the plant canopy part of the digital image (without the interference of the soil or water surface), and then the "histogram procedure" was employed to obtain data. The R, G, and B values of the image were measured, and the corresponding NRI, NGI, and NBI were calculated. The calculation of each normalized value is as follows:

$$\text{NRI} = R/(R + G + B)$$

$$\text{NGI} = G/(R + G + B)$$

$$\text{NBI} = B/(R + G + B)$$

3) Statistical analysis and model validation

The yield difference between treatments was statistically analyzed in SPSS16.0. Differences were compared with the Duncan method, with differences in N fertilizer application rate and planting density distinguished. When $p < 0.05$, the difference was significant.

The evaluation model was constructed through linear relationships between grain yield and color parameters through data of 2016. In order to test the reliability and universality of the model, the established models were validated using the data of 2017. The validity of the models was estimated from the statistical values of RMSE (root mean square error), RE (relative error), and RRMSE (relative root mean square error), which were calculated as:

$$\text{RMSE} = \sqrt{\frac{\sum (X_0 - X_S)^2}{n}} \quad (1)$$

$$\text{RE} = \frac{X_0 - X_S}{X_0} \times 100\% \quad (2)$$

$$\text{RRMSE} = \sqrt{\frac{1}{n} \sum \left(\frac{X_S - X_0}{X_0} \right)^2} \quad (3)$$

where X_0 and X_S represent measured and predicted values, respectively. The model is available when $\text{RRMSE} < 25\%$.

Additional file

Additional file 1: Fig. S1. The rice growth of tillering (A), jointing (B), heading (C), filling (D), late filling (E) and maturity (F) stages in 2016. **Fig. S2** The rice growth of late filling stage in 2017.

Abbreviations

R: red light; G: green light; B: blue light; N: nitrogen; NRI: normalized redness intensity; NGI: normalized greenness intensity; NBI: normalized blueness intensity; NDVI: normalized difference vegetation index; SPAD: soil-plant analyses development; SAVI: soil adjustment vegetation index; DGCI: dark green color index; LAI: leaf area index; GMR: green-channel minus red; RMSE: root mean square error; RE: relative error; RRMSE: relative root mean square error.

Authors' contributions

KL, YL, and TH designed the study; KL and TH measured rice yield and rice canopy. XY and HY managed the field experiment. KL, HH, and ZH conducted statistical analyses and drafted the manuscript. All authors contributed to the interpretation of results and/or drafting the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The data generated or analyzed during this study are included in this published article and its supplementary information files.

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Not applicable.

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References

- Wang W, Mauleon R, Hu Z, Chebotarov D, Tai S, Wu Z, Mansueto L. Genomic variation in 3010 diverse accessions of Asian cultivated rice. *Nature*. 2018;557:43–9.
- Khush GS. What it will take to feed 5.0 billion rice consumers in 2030. *Plant Mol Biol*. 2005;59(1):1–6.
- Xiao X, Boles S, Frohling S, Li C, Babu JY, Salas W, Moore B III. Mapping paddy rice agriculture in South and Southeast Asia using multi-temporal MODIS images. *Remote Sens Environ*. 2006;100(1):95–113.
- Muthayya S, Sugimoto JD, Montgomery S, Maberly GF. An overview of global rice production, supply, trade, and consumption. *Ann NY Acad Sci*. 2014;1324(1):7–14.
- Hansen PM, Schjoerring JK. Reflectance measurement of canopy biomass and nitrogen status in wheat crops using normalized difference vegetation indices and partial least squares regression. *Remote Sens Environ*. 2003;86(4):542–53.
- Son NT, Chen CF, Chen CR, Minh VQ, Trung NH. A comparative analysis of multitemporal MODIS EVI and NDVI data for large-scale rice yield estimation. *Agric Forest Meteorol*. 2014;197:52–64.
- Williams JD, Kitchen NR, Scharf PC, Stevens WE. Within-field nitrogen response in corn related to aerial photograph color. *Precis Agric*. 2010;11(3):291–305.
- Huang J, Wang X, Li X, Tian H, Pan Z. Remotely sensed rice yield prediction using multi-temporal NDVI data derived from NOAA's AVHRR. *PLoS ONE*. 2013;8(8):e70816.
- Liu KL, Li YZ, Hu HW. Predicting ratoon rice growth rhythm based on NDVI at key growth stages of main rice. *Chil J Agric Res*. 2015;75(4):410–7.
- Wang YP, Chang KW, Chen RK, Lo JC, Shen Y. Large-area rice yield forecasting using satellite imageries. *Int J Appl Earth Obs*. 2010;12(1):27–35.
- Li T, Hasegawa T, Yin X, Zhu Y, Boote K, Adam M, Gaydon D. Uncertainties in predicting rice yield by current crop models under a wide range of climatic conditions. *Global Change Biol*. 2015;21(3):1328–41.
- Hjorth L, Pink S. New visualities and the digital wayfarer: reconceptualizing camera phone photography and locative media. *Mobile Media Commun*. 2014;2(1):40–57.
- Jiang HY, Ying YB, Wang JP, Rao XQ, Xu HR, Wang MH. Real time intelligent inspecting and grading line of fruits. *Trans CSAE*. 2002;18(6):158–60 (in Chinese with English abstract).
- Dammer KH, Möller B, Rodemann B, Heppner D. Detection of head blight (*Fusarium* spp.) in winter wheat by color and multispectral image analyses. *Crop Prot*. 2011;30(4):420–8.
- Kawashima S, Nakatani M. An algorithm for estimating chlorophyll content in leaves using a video camera. *Ann Bot*. 1998;81(1):49–54.
- Adamsen FG, Pinter PJ, Barnes EM, LaMorte RL, Wall GW, Leavitt SW, Kimball BA. Measuring wheat senescence with a digital camera. *Crop Sci*. 1999;39(3):719–24.
- Rorie RL, Purcell LC, Mozaffari M, Karcher DE, King CA, Marsh MC, Longer DE. Association of "Greenness" in corn with yield and leaf nitrogen concentration. *Agron J*. 2011;103(2):529–35.
- Li Y, Chen D, Walker CN, Angus JF. Estimating the nitrogen status of crops using a digital camera. *Field Crop Res*. 2010;118(3):221–7.
- Lee KJ, Lee BW. Estimation of rice growth and nitrogen nutrition status using color digital camera image analysis. *Eur J Agron*. 2013;48:57–65.
- Wang Y, Wang D, Shi P, Omasa K. Estimating rice chlorophyll content and leaf nitrogen concentration with a digital still color camera under natural light. *Plant Method*. 2014;10(1):36.
- Wang Y, Wang D, Zhang G, Wang J. Estimating nitrogen status of rice using the image segmentation of GR thresholding method. *Field Crop Res*. 2013;149:33–9.
- Zhou X, Zhen HB, Xu XQ, He JY, Ge XK, Yao X, Tian YC. Predicting grain yield in rice using multi-temporal vegetation indices from UAV-based multispectral and digital imagery. *ISPRS J Photogramm Remote Sens*. 2017;130:246–55.
- Bian ZH, Yang QC, Liu WK. Effects of light quality on the accumulation of phytochemicals in vegetables produced in controlled environments: a review. *J Sci Food Agric*. 2015;95(5):69–877.
- Arena C, Tsonev T, Doneva D, De Micco V, Michelozzi M, Brunetti C, Loreto F. The effect of light quality on growth, photosynthesis, leaf anatomy and volatile isoprenoids of a monoterpene-emitting herbaceous species (*Solanum lycopersicum* L.) and an isoprene-emitting tree (*Platanus orientalis* L.). *Environ Exp Bot*. 2016;130:122–32.
- Long JR, Wan YZ, Song CF, Sun J, Qin RJ. Effects of nitrogen fertilizer level on chlorophyll fluorescence characteristics in flag leaf of super hybrid rice at late growth stage. *Rice Sci*. 2013;20(3):220–8.
- Havé M, Marmagne A, Chardon F, Masclaux-Daubresse C. Nitrogen remobilization during leaf senescence: lessons from *Arabidopsis* to crops. *J Exp Bot*. 2016;68(10):2513–29.

27. Kim J, Shon J, Lee CK, Yang W, Yoon Y, Yang WH, Lee BW. Relationship between grain filling duration and leaf senescence of temperate rice under high temperature. *Field Crop. Res.* 2011;122(3):207–13.
28. Yi R, Zhu Z, Hu J, Qian Q, Dai J, Ding Y. Identification and expression analysis of microRNAs at the grain filling stage in rice (*Oryza sativa* L.) via deep sequencing. *PLoS ONE.* 2013;8(3):e57863.
29. Dou F, Soriano J, Tabien RE, Chen K. Soil texture and cultivar effects on rice (*Oryza sativa*, L.) grain yield, yield components and water productivity in three water regimes. *PLoS ONE.* 2016;11(3):e0150549.
30. Chen S, Ge Q, Chu G, Xu C, Yan J, Zhang X, Wang D. Seasonal differences in the rice grain yield and nitrogen use efficiency response to seedling establishment methods in the middle and lower reaches of the Yangtze River in China. *Field Crop Res.* 2017;205:157–69.
31. Rosenzweig C, Iglesias A, Yang XB, Epstein PR, Chivian E. Climate change and extreme weather events; implications for food production, plant diseases, and pests. *Global Change Hum Health.* 2001;2(2):90–104.
32. Okuno A, Hirano K, Asano K, Takase W, Masuda R, Morinaka Y, Matsuoka M. New approach to increasing rice lodging resistance and biomass yield through the use of high gibberellin producing varieties. *PLoS ONE.* 2014;9(2):e86870.
33. Corbin JL, Orlowski JM, Harrell DL, Golden BR, Falconer L, Krutz LJ, Walker TW. Nitrogen strategy and seeding rate affect rice lodging, yield, and economic returns in the midsouthern United States. *Agron J.* 2016;108(5):1938–43.
34. Jia LL, Chen XP, Zhang FS, Buerkert A, Römheld V. Use of digital camera to assess nitrogen status of winter wheat in the northern China plain. *J Plant Nutr.* 2004;27(3):441–50.

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